



电膜萃取-液相色谱质谱法高通量检测尿液中内源性肽的研究*

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【摘要】目的 建立一种基于电膜萃取-液相色谱质谱联用技术精准检测尿中肽的方法,并评估其在肿瘤标志物筛选的应用前景。**方法** 选取15种与疾病相关的肽作为目标分析物,采用含有5%二(2-乙基己基)磷酸酯的正辛醇作为支撑液膜,供体相为100 mmol/L甲酸与尿液1:1混合液,受体相为20 mmol/L甲酸(含50%二甲基亚砜),40 V电压下电膜萃取15 min,受体相溶液经液相色谱串联质谱分析。初步将该方法应用于12例健康对照和7例泌尿系统肿瘤患者的尿样分析。**结果** 15种小分子肽在0.1~100.0 ng/mL范围内呈良好线性关系($r \geq 0.995$),检出限为0.01~0.50 ng/mL,定量限为0.03~1.50 ng/mL,加标回收率为21.0%~71.2%,相对标准偏差为0.8%~20.0%($n=3$)。小样本临床标本检测数据显示,肿瘤患者尿液中缓激肽1-5浓度(中位数0.65 ng/mL)高于健康对照组(中位数0.37 ng/mL),差异有统计学意义($P < 0.05$),有望成为泌尿系统肿瘤的特异性标志物。**结论** 本研究建立的电膜萃取-液相色谱质谱联用方法具有简便、高效、灵敏等优势,可实现尿液中痕量肽的精准检测,为疾病标志物筛查及电膜萃取技术的临床应用提供了可靠的方法学基础。

【关键词】 电膜萃取 微萃取 肿瘤标志物 尿液 肽

High-Throughput Determination of Endogenous Peptides in Urine Using Electromembrane Extraction Combined With Liquid Chromatography-Tandem Mass Spectrometry

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[Abstract] Objective To develop a precise method for analyzing urinary peptides based on electromembrane extraction (EME) combined with liquid chromatography-tandem mass spectrometry (LC-MS/MS), and to evaluate its potential applicability in tumor biomarker screening. **Methods** A total of 15 disease-associated peptides were selected as the target analytes. A supported liquid membrane (SLM) composed of n-octanol containing 5% di (2-ethylhexyl) phosphate was employed, with the donor phase being a 1:1 mixture of urine and 100 mmol/L formic acid and urine, and the acceptor phase being 20 mmol/L formic acid containing 50% dimethyl sulfoxide (DMSO). After EME at 40 V for 15 min, the acceptor phase solution was analyzed by LC-MS/MS. Subsequently, the method, EME combined with LC-MS/MS (EME-LC-MS/MS), was preliminarily validated utilizing urine samples from 12 healthy controls and 7 patients with urinary system tumors. **Results** All 15 peptides exhibited excellent linearity in the range of 0.1-100.0 ng/mL ($r \geq 0.995$), with the limits of detection (LODs) being 0.01-0.50 ng/mL and the limits of quantification (LOQs) being 0.03-1.50 ng/mL. The spiked recoveries ranged from 21.0% to 71.2%, with relative standard deviations (RSDs) of 0.8%-20.0% ($n = 3$). Small-sample analysis of clinical specimens revealed that the concentration of bradykinin 1-5 in the urine were significantly higher in tumor patients (median: 0.65 ng/mL) than that in healthy controls (median: 0.37 ng/mL) ($P < 0.05$), suggesting its potential as a specific biomarker for urinary system tumors. **Conclusion** The EME-LC-MS/MS method established in the study features simplicity, high efficiency, and high sensitivity, enabling precise determination of trace-level peptides in urine samples. Moreover, this approach provides a reliable methodological basis for disease biomarker screening and promotes the clinical application of electromembrane extraction.

[Key words] Electromembrane extraction Microextraction Tumor biomarkers Urine Peptides

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恶性肿瘤是目前全球公共卫生领域的重大挑战,中国约占全球新发癌症病例的25%^[1-2]。据预测,2050年中国的癌症新发病例会较2022年增长51.4%^[3]。近年来,基于肿瘤标志物的无创检测技术开发逐渐成为研究热点。有研

究发现,小分子肽参与多种生理病理进程的调控,可反映机体病理生理状态^[4-5]。尿液中的小分子肽和慢性肾病^[6]、系统性红斑狼疮^[7]、慢性疼痛^[8],以及肾移植预后^[9]等相关,可望成为新一代疾病标志物。但尿液具有高盐高尿素的复杂基质,且尿液中内源性小分子肽丰度较低^[10]。亟需开发简便、高通量的前处理技术,用于高效分离富集尿液中小分子肽,并联用液相色谱串联质谱技术实现精准检测。

固相萃取是目前小分子肽分析中最常用的前处理技术,具有高选择性、操作自动化等优势,但其有机溶剂使用量高,难以满足绿色化学要求^[11]。微萃取技术凭借其低溶剂消耗、高富集能力和操作简便等优势,逐渐成为前处理技术的研究热点^[12]。2006年, PEDERSEN-BJERGAARD 教授^[13]首次提出电膜萃取(electromembrane extraction, EME)技术,该技术通过电场驱动带电目标物选择性跨膜迁移,从而实现目标物的定向分离与富集,具有绿色、高效等优点,可为复杂生物样本中痕量物质的富集提供创新解决方案。随后,EME作为一种新型的绿色微萃取技术被广泛应用于生物材料检验、环境监测等领域^[14-16]。

因此,本研究针对传统前处理技术中基质净化能力和分析灵敏度不足的瓶颈,创新性地构建了96孔电膜萃取-液相色谱串联质谱分析平台,用于尿液中15种特征小分子肽的高通量提取与精准检测,并初步应用于真实尿样的分析,可为肿瘤无创筛查提供高稳定性、高效的分析工具。

1 材料与方法

1.1 仪器与设备

高效液相色谱仪(日本, SHIMADZU)串联三重四级

杆质谱(美国, AB SCIEX 6500)系统;紫外-可见分光光度计(日本, SHIMADZU UV-2700i);数控圆周摇床(中国, DLAB SK-O180-Pro);可编程线性直流电源(中国, INTERLOCK IPD200-1A);万用表(美国, FLUKE 287 C);实验室自主设计96孔电膜萃取装置。

1.2 试剂与材料

15种目标肽(分子质量范围为300~1700,净电荷量为0.77~9.85)的纯度及相关理化参数详见表1(数据来源:chemicalize.com),谷氨酰-谷氨酰-亮氨酸(Glu-Glu-Leu)、促性腺激素释放激素(gonadotropin-releasing hormone, GnRH)购自中国丹港生物科技公司,蛋氨酸脑啡肽(met-enkephalin)、血管紧张素1-7(angiotensin 1-7)、血管紧张素II(3-8)[cangiotensin II(3-8)]、血管紧张素II(angiotensin II)、血管紧张素I(angiotensin I)、去精氨酸(9)缓激肽[Des-Arg(9)-bradykinin]、缓激肽1-7(bradykinin 1-7)、缓激肽1-5(bradykinin 1-5)购自中国麦克林公司,亮氨酸脑啡肽(leu-enkephalin)、神经降压肽(neurotensin)、P物质(neurokinin P)、缓激肽(bradykinin)购自美国AbMole公司,缓激肽2-9(bradykinin 2-9)购自美国MedChemExpress公司。支撑液膜溶剂包括:2-硝基苯辛醚(2-nitrophenyl octyl ether, NPOE)、2-十一酮(2-undecanone)、香芹酚(carvacrol)、磷酸三正戊酯(tripentyl phosphate, TAMP)、二(2-乙基己基)磷酸酯(di(2-ethylhexyl) phosphate, DEHP)、三(2-乙基己基)磷酸酯(tris(2-ethylhexyl) phosphate, TEHP)、正辛醇(1-Octanol)、1-乙基-2-硝基苯(1-ethyl-2-nitrobenzene, ENB)均购自阿拉丁公司;亚磷酸双(2-乙基己基)酯[Bis(2-ethylhexyl) phosphite, DEHPi]购自麦克林公司。

表1 肽的物理化学参数和质谱参数

Table 1 The physicochemical and mass spectrometry parameters of peptides

Peptide name	Sequence	Purity	pKa	Z (pH 3.0)	Molecular mass	RT/min	Q1	Q3	DP/V	CE/V
Glu-Glu-Leu	EEL	98%	3.35	0.77	389.18	1.06	390.1	132.0 [#] /86.0	60/60	19/42
Met-enkephalin	YGGFM	> 99%	3.91	1.31	573.22	1.97	574.4	425.0 [#] /296.9	55/55	23/30
Leu-enkephalin	YGGFL	> 99%	4.00	1.33	555.27	2.22	556.1	396.9 [#] /538.0	40/40	29/22
Neurotensin	pE-LYENKPRRPYIL	> 97%	3.65	8.83	1671.91	1.94	558.5	643.3 [#] /578.9	41/39	32/30
Gonadotropin-releasing hormone	pE-HWSYGLRPG	98%	-2.90	9.85	1182.56	1.87	592.6	499.2 [#] /935.2	39/58	29/30
Neurokinin P	RPKPQQFFGLM	> 99%	10.15	7.32	1346.73	2.15	674.7	600.4 [#] /586.3	63/40	31/34
Angiotensin 1-7	DRVYIHP	> 98%	3.35	5.19	898.47	1.30	450.5	392.7 [#] /647.1	47/44	20/25
Angiotensin II (3-8)	VYIHPF	98%	3.90	3.95	774.41	2.05	775.4	513.1 [#] /485.4	80/80	38/42
Angiotensin II	DRVYIHPF	90%	3.57	5.46	1045.53	1.97	524.1	263.1 [#] /784.3	70/70	28/27
Angiotensin I	DRVYIHPFHL	> 98%	3.50	7.57	1295.67	2.15	433.2	513.0 [#] /647.0	62/55	20/21
Bradykinin	RPPGFSPFR	> 99%	3.88	5.78	1059.56	1.73	531.1	522.0 [#] /807.0	67/78	30/36
Des-Arg (9)-bradykinin	RPPGFSPF	95%	3.81	4.55	903.46	2.08	452.8	642.1 [#] /262.8	33/33	21/20
Bradykinin 1-7	RPPGFSP	> 97%	3.43	4.09	756.39	1.34	379.3	555.4 [#] /641.8	23/45	18/16
Bradykinin 2-9	PPGFSPFR	97%	3.81	4.55	903.46	1.94	452.9	404.0 [#] /710.0	69/65	25/29
Bradykinin 1-5	RPPGF	> 97%	3.45	3.43	572.30	0.94	573.3	320.1 [#] /236.8	57/27	47/43

RT: retention time; Q1: precursor ion; Q3: product ion; DP: declustering potential; CE: collision energy. [#] Quantitative ion pair.

二甲基亚砜(dimethyl sulfoxide, DMSO)购自德国BioFroxx公司。质谱纯的甲酸(formic acid, FA)和甲醇购自美国Fisher公司。0.45 μm 孔径的PVDF 96孔滤板购于美国Millipore公司。实验用水为超纯水(电阻率为18.25 $\text{M}\Omega\cdot\text{cm}$)。尿样由四川省肿瘤医院提供(伦理批号:SCCHEC-02-2023-016),所有尿样均为空腹8 h以上中段晨尿,采集于15 mL无菌离心管中并于采集后2 h内置于 $-80\text{ }^{\circ}\text{C}$ 保存。

1.3 溶液配制

将各目标肽溶于体积分数30%的甲醇水溶液,配制1.0 mg/mL单标储备液。混合单标溶液得到15种肽的混合储备液(10.0 $\mu\text{g}/\text{mL}$)。临用前用样品基质(含甲酸与尿

液的稀释液)稀释至终浓度为100.0 ng/mL的工作液。采用受体相溶液配制0.1、0.2、0.5、1.0、2.0、5.0、10.0、20.0、50.0、100.0 ng/mL的标准系列浓度。

1.4 电膜萃取程序

本研究采用自主设计的96孔电膜萃取装置(图1)。取3 μL 正辛醇(含5% DEHP)均匀滴加于PVDF膜表面作为支撑液膜(supported liquid membrane, SLM);取250 μL 标准溶液或尿液甲酸1:1稀释液加入金属孔中作为供体相;取50 μL 受体相溶液(含50% DMSO的20 mmol/L 甲酸)于96孔滤板中作为受体相,此为一个提取单元。40 V恒压条件下电膜萃取15 min,实时监测电流。收集受体相溶液进行LC-MS/MS分析。

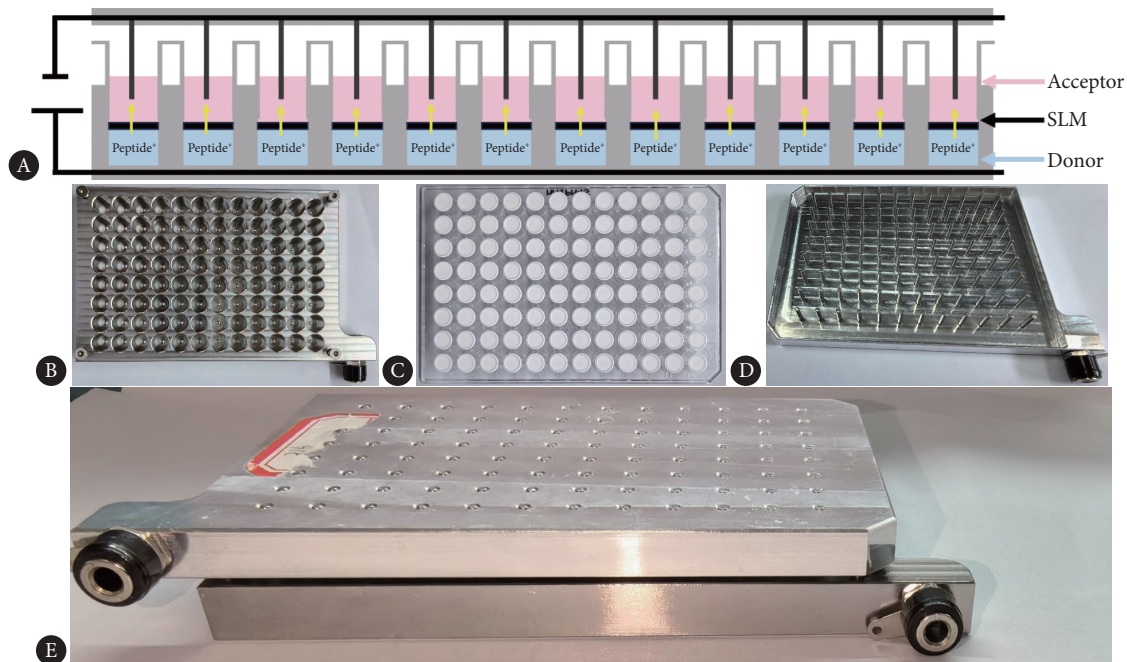


图1 电膜萃取原理图及实物图

Fig 1 Schematic illustration and photographs of the electromembrane extraction device

SLM: supported liquid membrane. A, Schematic of electromembrane extraction; B, metal well plate; C, polyvinylidene fluoride (PVDF) filter plate; D, lid with rod-like electrodes; E, a 96-well electromembrane extraction device already assembled.

1.5 LC-MS/MS分析

采用Waters ACQUITY UPLC BEH C_{18} 色谱柱(2.1 mm \times 50 mm, 2.5 μm)分离受体相溶液,流动相 A(5%甲醇水溶液,含0.1%的甲酸)和流动相 B(95%甲醇水溶液,含0.1%的甲酸)进行梯度洗脱,洗脱程序:0~2 min, 20%~80% B, 2~3 min, 80% B, 3~4 min, 20% B。流速0.3 mL/min,柱温40 $^{\circ}\text{C}$,进样量5 μL 。根据保留时间和质荷比(m/z)定性,峰面积定量。

采用电喷雾离子源(ESI^+),多反应监测模式(MRM)。离子化电压5 500 V,离子源温度600 $^{\circ}\text{C}$,气帘气35 psi,碰撞气8 psi,喷雾气60 psi,辅助加热气70 psi。目

标物的质谱参数见表1。

1.6 方法学考察

采用标准曲线法评估方法的定量性能,测定0.1~100.0 ng/mL系列浓度标准溶液,通过加权最小二乘法建立校准曲线。以3倍和10倍信噪比(S/N)对应的浓度分别作为方法的检出限和定量限。采用加标回收实验评估方法的准确度和精密度。

1.7 统计学方法

采用GraphPad Prism 9.1.0软件进行统计学分析和绘图。为比较病例组和对照组之间的差异,连续变量经正态性检验,若满足正态分布且方差齐性,两组间比较采用

独立样本 *t* 检验; 若不满足正态分布或方差不齐, 则采用 Wilcoxon 秩和检验。 *P* < 0.05 为差异有统计学意义。

2 结果

2.1 支撑液膜的优化

在电膜萃取过程中, SLM 的溶剂化能力是驱动带电分析物跨膜传质的关键机制, 其效率依赖于溶质与液膜间的氢键、 π - π 键及离子相互作用。基于此, 本研究考察了 2-硝基苯辛醚、2-十一酮、磷酸三正戊酯、香芹酚、正

辛醇等常见有机溶剂的萃取性能。结果(图2A)显示, 单一有机溶剂对亲水性肽的回收率普遍低于20%, 传统液膜难以有效富集极性多肽。为提升传质效率, 研究进一步选择正辛醇作为基础支撑液膜, 并引入阴离子载体二(2-乙基己基)磷酸酯(DEHP), 通过形成离子对络合物增强质子化肽的跨膜迁移能力。实验发现, 正辛醇提取目标肽的平均回收率仅为14.5%; 当添加2% DEHP后, 回收率显著提升至27.3%, 证实离子交换机制对肽类富集的显著促进作用。

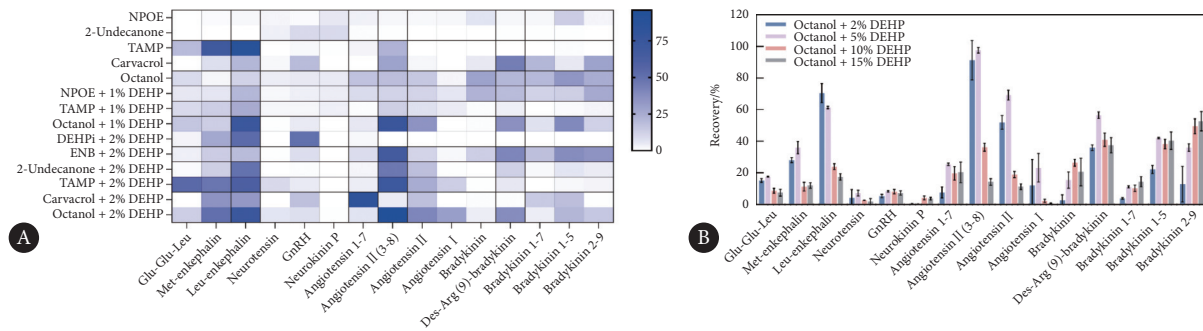


图 2 不同 SLM (A) 和正辛醇中添加不同浓度 DEHP (B) 对目标物的提取性能影响 (n=3)

Fig 2 The effect of different supported liquid membranes (A) and the addition of DEHP at different concentrations (B) in n-octanol on the performance of the extraction of target substances (n = 3)

离子载体的添加量不足时, 传质驱动力减弱; 而浓度过高时, 会导致支撑液膜电导率升高从而引发电解副反应(如 pH 偏移或气泡生成), 降低体系稳定性。本实验进一步系统考察了 DEHP 添加比例(2% ~ 15%)对萃取效率的影响。结果表明(图2B), 当 DEHP 浓度低于 5% 时, 液膜离子交换容量不足; 当 DEHP 浓度升高至 5% 时, 提取效率达到峰值(平均回收率 33.9%); 当 DEHP 浓度超过 10% 时, 液膜导电性显著升高, 焦耳热效应加剧, 导致稳定性下降, 回收率出现降低趋势。综上, 选择正辛醇(含 5% DEHP)作为最优支撑液膜, 可兼具高传质效率(平均回收率 > 30%)和系统稳定性。

2.2 供体相与受体相 pH 值的协同优化

电膜萃取过程中, 目标物的质子化状态直接决定其有效电荷量及电迁移速率。本研究通过调控供体相与受体相溶液的 pH 值(2.2 ~ 2.9), 系统探究了受体相和供体相的 pH 值对 15 种肽提取性能的影响。如图 3 所示, 血管紧张素衍生物及亮氨酸脑啡肽等碱性氨基酸肽在低 pH 区间(pH 2.2 ~ 2.5)时, 由于氨基质子化增强, 回收率随 pH 值的增大而提升。缓激肽系列物质在高 pH 条件(pH 2.5 ~ 2.9)下因羧基质子化抑制, 回收率提高了约 2.5 倍。值得注意的是, 当 pH > 2.9 时, 部分肽因去质子化导致净电荷趋近中性, 电迁移驱动力丧失, 回收率骤降。同时考虑到

高浓度甲酸缓冲体系虽可维持强酸性环境, 但会引发电解产气现象, 破坏液膜的稳定性。基于多目标物协同富集需求, 最终选定两相的 pH 值均为 2.9, 可满足不同分析物质子化需求且有效抑制过度电解引发的副反应。

2.3 改性剂浓度的优化

在电膜萃取体系中, DMSO 可作为受体相改性剂调节液膜界面化学微环境, 从而改善 $pK_a < 4$ 的分析物在液膜上的传质效率^[17]。因此, 本研究系统考察受体相中含有不同含量 DMSO(10%、25%、50%、75%) 时 15 种低

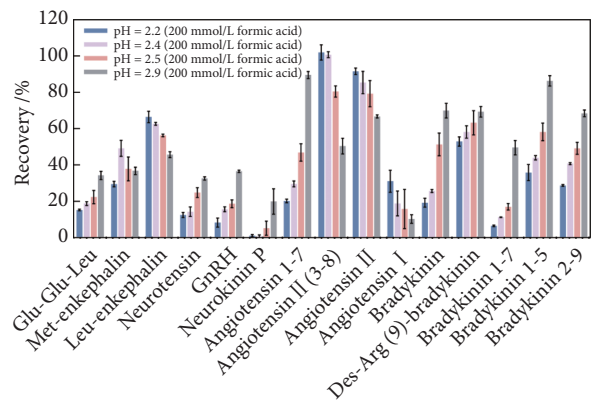


图 3 不同两相 pH 值对目标物的提取效率影响 (n=3)

Fig 3 Effect of different pH values of donor and acceptor phases on the extraction efficiency of target substances (n = 3)

pKa肽(pKa -2.90 ~ 10.15)电膜提取性能。结果如图4所示,当DMSO含量从10%增至50%时,Glu-Glu-Leu、血管紧张素衍生物等含酸性肽回收率提高了2.1~3.8倍,归因于DMSO可抑制羧酸根与液膜阴离子载体的静电吸附。脑啡肽类及缓激肽类肽,在含50% DMSO的受体相时,回收率分别达到73.8%~83.8%和32.9%~88.6%,较10% DMSO受体相体系提高2倍以上;当DMSO浓度进一步升高至75%,缓激肽代谢物1-7、1-5和2-9的回收率略降,可能是受体相黏度显著增加导致的传质阻力增大而引起回收率的微降。因此,为维持受体相质子化能力,本研究选择20 mmol/L甲酸(含50% DMSO, pH≈2.9)作为最佳受体相。

2.4 线性范围与灵敏度

如表2所示,15种目标肽在线性范围内呈现良好线性,相关系数(r) ≥ 0.995 。方法检出限(limits of detection, LODs)与定量限(limit of quantitation, LOQs)分别为0.01~0.50 ng/mL和0.03~1.50 ng/mL,满足小分子肽的痕量分析要求。

2.5 基质效应、加标回收率和稳定性

如图5所示,目标物在尿液基质与缓冲溶液中的提取率分别在10.0%~58.9%和21.1%~79.9%。基质效应(matrix effects, ME)为71.8%~150.7%,其中血管紧张素

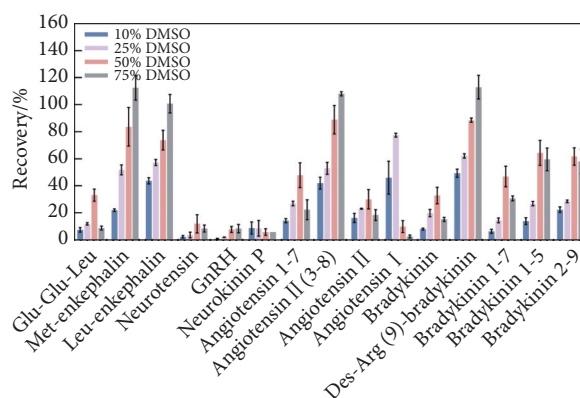


图4 受体相中改性剂含量对目标物的提取效率影响 ($n=3$)

Fig 4 Effect of different concentrations of dimethyl sulfoxide, a modifier, in the acceptor phase on the extraction efficiency of target substances ($n=3$)

II(3-8)的ME为150.7%,后续将继续优化实验条件,以降低该物质的基质效应。其余物质基质效应均满足分析要求,表明本研究建立的电膜萃取技术可有效去除尿液样本中基质的干扰。随后,测定3个加标浓度(5.0、10.0、15.0 ng/mL)的加标尿样。结果如表3所示,目标物的回收率在21.0%~71.2%,相对标准偏差为0.8%~20.0%,满足生物分析指南[欧洲药品管理局(EMA)]对痕量分析的精度要求。

表2 本方法的线性范围、灵敏度和稳定性 ($n=6$)

Table 2 Linear range, sensitivity, and stability of the established method ($n=6$)

Peptide	Linearity/(ng/mL)	Regression equation	r	LOD/(ng/mL)	LOQ/(ng/mL)	RSD/%	
						Intra-day	Inter-day
Glu-Glu-Leu	0.1-100.0	$y = 7.4 \times 10^5 x - 2.2 \times 10^4$	0.999	0.01	0.03	9.0	7.0
Met-enkephalin	0.1-100.0	$y = 3.4 \times 10^4 x + 590$	0.999	0.01	0.03	6.6	1.9
Leu-enkephalin	0.1-100.0	$y = 4.8 \times 10^5 x + 1.8 \times 10^5$	0.999	0.01	0.03	1.7	2.1
Neurotensin	2.0-100.0	$y = 1.1 \times 10^5 x - 1.6 \times 10^5$	0.995	0.01	0.03	3.6	0.6
Gonadotropin-releasing hormone	0.1-100.0	$y = 1.9 \times 10^5 x - 1.4 \times 10^4$	0.998	0.01	0.03	6.2	4.5
Neurokinin P	2.0-100.0	$y = 7.4 \times 10^3 x - 1.4 \times 10^4$	0.997	0.50	1.50	0.9	5.0
Angiotensin 1-7	0.5-100.0	$y = 2.2 \times 10^5 x - 7.6 \times 10^4$	0.999	0.02	0.06	5.2	2.7
Angiotensin II (3-8)	0.2-100.0	$y = 5.5 \times 10^4 x - 9.8 \times 10^3$	0.999	0.01	0.03	8.6	8.3
Angiotensin II	1.0-100.0	$y = 6.8 \times 10^5 x - 5.9 \times 10^5$	0.997	0.01	0.03	5.3	13.0
Angiotensin I	2.0-100.0	$y = 8.9 \times 10^3 x - 1.6 \times 10^4$	0.999	0.10	0.30	3.4	13.4
Bradykinin	2.0-100.0	$y = 6.1 \times 10^4 x - 1.1 \times 10^5$	0.999	0.05	0.15	4.0	8.5
Des-Arg (9)-bradykinin	0.5-100.0	$y = 3.6 \times 10^5 x - 1.7 \times 10^5$	0.999	0.01	0.03	0.8	6.3
Bradykinin 1-7	0.5-100.0	$y = 1.0 \times 10^5 x - 2.8 \times 10^4$	0.999	0.01	0.03	2.9	3.5
Bradykinin 1-5	0.1-100.0	$y = 9.6 \times 10^4 x - 2.1 \times 10^4$	0.999	0.02	0.06	0.5	3.2
Bradykinin 2-9	0.1-100.0	$y = 3.5 \times 10^5 x + 1.9 \times 10^4$	0.999	0.01	0.03	3.9	4.8

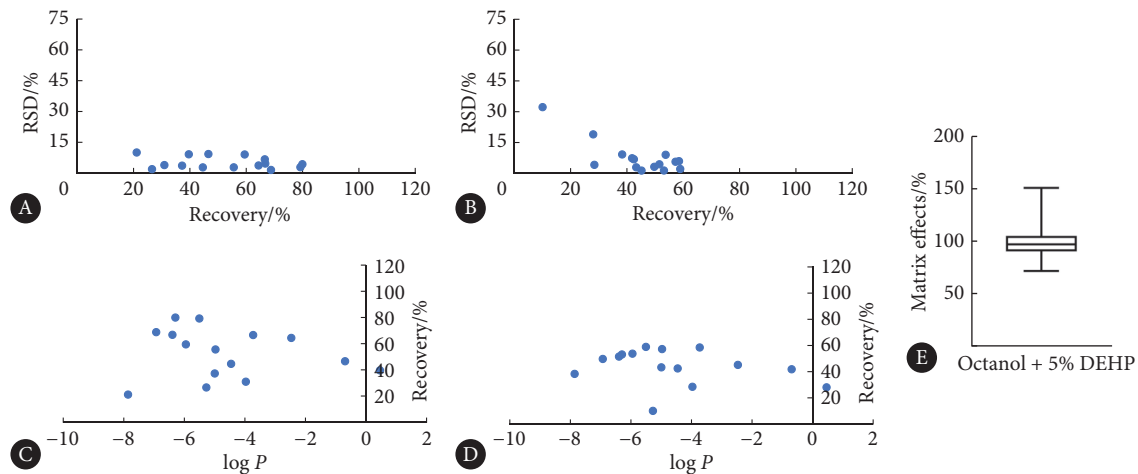


图5 加标回收率和基质效应考察

Fig 5 Evaluation of spike and recovery and matrix effects

A and C, Extraction performances of peptides in the buffer solution; B and D, extraction performances of peptides in the urine sample; E, results of matrix effect experiments.

2.6 尿液样本的初步应用

为评估方法的临床适用性,采用本研究构建的电膜萃取-液相色谱质谱联用技术测定了4例膀胱癌患者和3例肾癌患者尿样,以及12例匹配了年龄/性别的健康对照组的晨尿样本。如图6所示,肿瘤患者尿中缓激肽1-5水平(0.37 ~ 2.45 ng/mL,中位数0.65 ng/mL)高于对照组(0.31 ~ 0.80 ng/mL,中位数0.37 ng/mL),经Wilcoxon秩和检验差异有统计学意义($P < 0.05$)。本研究结果提示,尿中缓激肽1-5的精准分析可望用于肿瘤早期非侵入式筛查和诊断。

3 讨论

本研究成功构建了基于96孔电膜萃取-液相色谱串联质谱技术平台,通过优化电膜萃取关键实验参数,提高了亲水性肽的跨膜传质效率,最终以含5%二(2-乙基己基)磷酸酯的正辛醇作为支撑液膜,供体相为100 mmol/L甲酸与尿液1:1混合液,受体相为20 mmol/L甲酸(含50%二甲基亚砜),40 V电压下电膜萃取15 min,实现尿中15种小分子肽的高效分离富集与检测。为揭示支撑液膜的作用机制,本研究进一步采用溶剂法^[18]测定了正辛醇(含5% DEHP)的Kamlet-Taft参数,氢键供体能力(α)、氢键受体能力(β)及极性/极化率(π^*)值分别为0.94、1.76和0.11。高 β 值表明液膜具有强氢键受体特性,可与肽氨基形成定向作用;同时,DEHP的磷酸酯基团通过离子交换可促进质子化肽的跨膜迁移。氢键-离子交换协同机制显著提高了亲水性肽的传质效率,符合离子载体增强两相界面传质的理论模型^[19]。本方法通过整合电场驱动传

质与质谱检测,首次实现尿源性肽的痕量定量,该方法线性范围宽,精密度良好,灵敏度较ELISA法提升超10倍,满足痕量生物标志物分析要求。

此外,本研究采用的96孔电膜萃取装置不仅兼具环境友好与高通量优势,而且可与质谱平台良好衔接,该方法与传统前处理方法比较(表4),EME作为一种绿色微萃取技术,有机溶剂消耗仅需3 μ L,远低于传统固相萃取方法(通常大于1 mL),前处理时间缩短至15 min(传统方法常需数小时),且无须额外富集浓缩步骤,操作简便,为高通量临床样本分析提供了高效且环保的解决方案。

本研究初步临床应用发现,泌尿系统肿瘤患者尿液中缓激肽1-5浓度高于健康对照组。有基础研究发现^[24-25],缓激肽1-5可通过竞争性抑制凝血酶活性,降低纤维蛋白原裂解效率,从而使血液可能呈现低凝状态。本研究在泌尿系统肿瘤患者尿中检出较高浓度的缓激肽1-5,可能与泌尿系统肿瘤患者血液常呈现低凝状态相关^[26],进一步证实小分子肽在泌尿肿瘤中具有较好的诊断潜力。然而,当前研究样本量较小,该标志物特异性、敏感性及其在肿瘤发生发展中的确切机制仍需通过大样本多中心研究并结合蛋白质组学/代谢组学技术系统解析其调控网络,进一步验证该标志物效能。综上所述,本研究可为尿液肽组学分析及肿瘤非侵入式筛查提供新的手段,其模块化设计也可扩展至其他生物标志物检测领域。

* * *

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表 3 尿中 15 种小分子肽的加标回收实验结果 ($n=3$)
 Table 3 The extraction performance of spiked experiments for 15 peptides in urine ($n = 3$)

Peptide	Background/(ng/mL)	Spiked/(ng/mL)	Found/(ng/mL)	Recovery/%	RSD/%
Glu-Glu-Leu	ND	5.0	1.0	21.0	8.2
		10.0	2.4	23.9	8.5
		15.0	4.0	26.5	4.9
Met-enkephalin	ND	5.0	3.6	71.2	10.7
		10.0	6.3	62.8	7.6
		15.0	9.6	63.7	1.0
Leu-enkephalin	ND	5.0	3.0	60.0	4.1
		10.0	6.5	65.0	2.8
		15.0	9.7	64.9	0.8
Neurotensin	1.45	5.0	2.5	50.3	7.4
		10.0	5.2	52.4	9.7
		15.0	7.5	50.3	0.8
Gonadotropin-releasing hormone	ND	5.0	2.1	42.4	1.8
		10.0	4.8	48.2	7.4
		15.0	7.1	47.1	2.4
Neurokinin P	1.94	5.0	2.9	57.3	8.8
		10.0	3.3	32.5	4.6
		15.0	4.7	31.4	20.0
Angiotensin 1-7	0.35	5.0	2.2	44.1	10.7
		10.0	5.5	55.3	16.1
		15.0	7.5	49.7	4.7
Angiotensin II (3-8)	0.19	5.0	2.4	47.3	10.7
		10.0	5.5	54.5	11.4
		15.0	8.5	56.3	4.4
Angiotensin II	0.87	5.0	2.1	41.2	2.1
		10.0	5.1	50.7	14.3
		15.0	7.5	49.7	2.1
Angiotensin I	1.87	5.0	1.9	37.3	16.2
		10.0	2.3	22.6	14.0
		15.0	4.4	29.6	19.6
Bradykinin	1.78	5.0	2.7	54.6	6.0
		10.0	6.2	62.0	14.9
		15.0	8.0	53.0	12.4
Des-Arg(9)-bradykinin	0.49	5.0	2.7	53.8	4.7
		10.0	6.3	63.1	11.7
		15.0	9.2	61.4	3.6
Bradykinin 1-7	0.29	5.0	1.4	28.3	2.2
		10.0	4.4	44.2	15.0
		15.0	8.2	54.8	6.5
Bradykinin 1-5	0.26	5.0	2.0	40.2	1.6
		10.0	5.4	54.5	12.2
		15.0	8.7	58.3	1.2
Bradykinin 2-9	ND	5.0	1.9	38.8	4.0
		10.0	6.0	60.2	10.1
		15.0	9.2	61.1	2.4

ND: not detected.

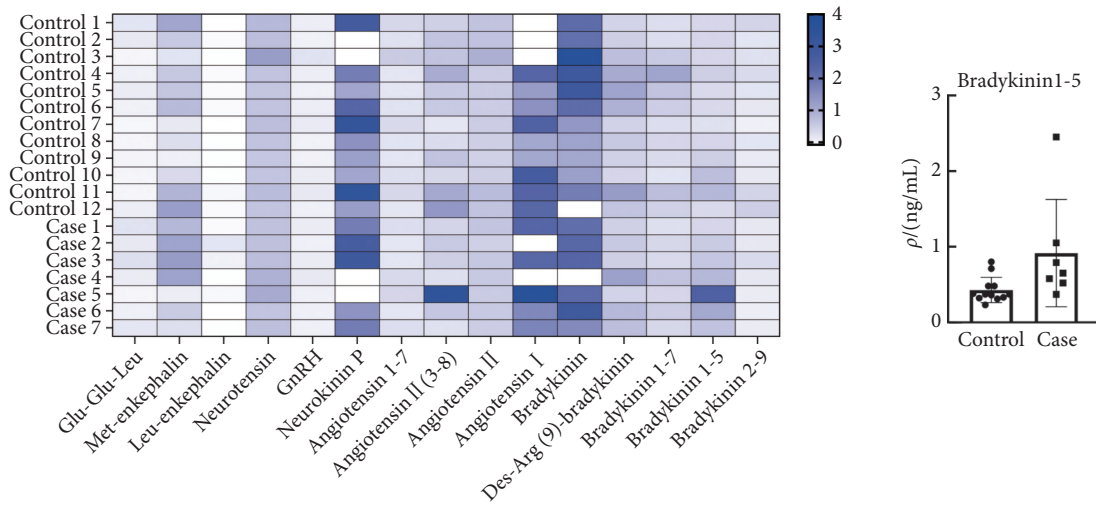


图 6 19份尿液标本中15种肽的浓度水平

Fig 6 Concentrations of 15 peptides in 19 urine specimens

表 4 与其他检测方法的比较

Table 4 Comparison with other methods

Pretreatment method	Organic solvent consumption	Extraction time	Number of analytes	Sample	Recovery/%	Ref
EME	3 μ L	15 min	15	Urine	21.0-71.2	This work
SPE-concentration	> 12 mL	> 2 h	9	Plasma, brain tissues	59.2-111.8	[20]
SPE-concentration	> 3 mL	> 30 min	4	Plasma	58.0-62.0	[11]
SPE-concentration	> 1 mL	> 4 h	7	Plasma	60.9-98.8	[21]
PP-concentration	265 μ L	16 min	14	Plasma	64.9-117.7	[22]
SPE	> 1 mL	> 4 h	3	plasma	81.0-113.0	[23]

EME: electromembrane extraction; SPE: solid phase extraction; PP: protein precipitation.

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